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## A Highly Efficient Copper(II) Complex catalysed Hydrolysis of Methyl Acetate at pH 7.0 and 25 °C

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The turnover time for  $[(2,2'-dipyridylamine)Cu(OH_2)_2]^{2+}$  (1 mm) catalysed hydrolysis of methyl acetate (1 m) is 23 min at pH 7, 25 °C.

Successful catalysed hydrolysis of activated esters is no guarantee that the same catalyst will hydrolyse unactivated esters.<sup>1</sup> We recently reported that a 10 mm solution of (1) gives a six-fold rate enhancement for methyl trifluoroacetate hydrolysis but no rate enhancement for methyl acetate

hydrolysis.<sup>2</sup> Indeed, true catalytic hydrolysis of unactivated esters under mild conditions has only been obtained with real enzymes despite enormous efforts to design efficient artificial esterases.<sup>3,4</sup> Here we report on efficient catalytic hydrolysis of methyl acetate using a simple Cu<sup>II</sup> complex (2).

A solution of (2) was standardised by titration with standard NaOH solution. The  $pK_a$  of the copper co-ordinated water molecule is 7.2 at 25 °C. Catalysed hydrolysis of methyl acetate (1 M) with (2) (0.3 to 1 mM) was monitored by the pH stat method.† The pH of the reaction solution was maintained with a Radiometer PHM63 pH meter equipped with a Radiometer RTS 822 automatic titrator. The catalytic turnover‡ time is 23 min at pH 7.0, 25 °C (Figure 1).

Based on the  $pK_a$  of the copper co-ordinated water molecule and the pH-rate profile (Figure 2), we propose that the mechanism of catalysed hydrolysis of methyl acetate using (2) involves co-ordination of the ester to the copper followed by intramolecular metal hydroxide attack on the co-ordinated ester (Scheme 1).<sup>5</sup> Since Cu<sup>II</sup> is substitutionally labile, either formation or breakdown of the tetrahedral intermediate is the rate-determining step ( $k_2$ ). The rate of acetic acid production is given by  $k_{obs}[(2)]_T$ [ester] where  $[(2)]_T$  is the total catalyst concentration and  $k_{obs}$  is given by equation (1). The pH-rate profile (Figure 2) was fitted according to equation (1) (Scheme 1).§ Under our experimental conditions, mono-aquo complexes such as (1) or (3) do not catalyse the hydrolysis of methyl acetate to any observable extent.

$$k_{\rm obs} = k_2 K_1 [K_a / (K_a + [\rm H^+])]$$
(1)

We chose to use 2,2'-dipyridylamine for its strong affinity towards Cu<sup>II</sup>. The ligand binds Cu<sup>II</sup> more tightly ( $K = 1.15 \times 10^8 \text{ mol}^{-1} \text{ dm}^3$ )<sup>6</sup> than it binds H<sup>+</sup> ( $K = 1.38 \times 10^7 \text{ mol}^{-1} \text{ dm}^3$ ).<sup>6</sup> Consequently, the metal ion does not dissociate from 2,2'-dipyridylamine over a wide range of the solution pH, including the p $K_a$  region for the copper-bound water molecule.

The second order rate constants  $(k_{obs})$  for catalysed hydrolysis of methyl acetate and *p*-nitrophenyl acetate using



<sup>†</sup> Methanol production was confirmed by <sup>1</sup>H n.m.r.

‡ At 1 mm catalyst concentration, one catalytic turnover every 23 min translates to a reaction rate of  $7.2 \times 10^{-7}$  mol<sup>-1</sup> dm<sup>3</sup> acetic acid produced per second  $[10^{-3}/(23 \times 60)]$ . The rate constants were reproducible to within 3%.

§ A non-linear least square curve fitting program was used to fit the data ( $K_a = 2.8 \times 10^{-7}$ ,  $K_1k_2 = 1.0 \times 10^{-3} \text{ mol}^{-1} \text{ dm}^3 \text{ s}^{-1}$ ). The  $K_a$  value obtained through potentiometric titration ( $6.3 \times 10^{-8}$ ) is more reliable than the one obtained kinetically.

(2) are  $7.2 \times 10^{-4}$  and  $1.6 \times 10^{-1}$  mol<sup>-1</sup> dm<sup>3</sup> s<sup>-1</sup> respectively (pH 7.0, 25 °C). The rates for methyl acetate<sup>7</sup> and *p*-nitrophenyl acetate<sup>8</sup> hydrolyses with water are  $3 \times 10^{-10}$  and  $6 \times 10^{-7}$  s<sup>-1</sup> respectively. Therefore, (2) gives a greater rate acceleration for methyl acetate hydrolysis than for *p*-nitrophenyl acetate hydrolysis. Simple catalysts that are highly efficient at hydrolysing unactivated esters are not necessarily efficient at hydrolysing unactivated esters. For example, (1), (3) or imidazole gives a much greater rate acceleration for *p*-nitrophenyl acetate hydrolysis than for methyl acetate hydrolysis.<sup>1</sup>

The equilibrium constant  $(K_1, \text{ Scheme 1})$  for complexation of methyl acetate to the copper complex cannot be measured directly. However  $K_1$  can be approximated as follows. There is a linear free energy relationship between the basicity of the ligands (L) and the equilibrium constant for complexation of L to aqueous Cu<sup>II</sup> [equation (2)],<sup>9</sup> where  $K = [(H_2O)_5(Cu)L]^{2+/2}$  $[(Cu)(H_2O)_6]^{2+}[L]$  and  $pK_a$  is the acid dissociation constant for the conjugate acid of L. The  $pK_a$  of protonated methyl acetate is about  $-6.0.^{10}$  Therefore, the equilibrium constant for binding methyl acetate to aqueous Cu<sup>II</sup> should be about 2.6  $\times$  10<sup>-3</sup> mol<sup>-1</sup> dm<sup>3</sup> [equation (2)]. This is an extended extrapolation considering that equation (2) is based on a series of substituted pyridines. However, log K for  $L = H_2O$ calculated from equation (2)  $\log K = 0.45(-1.72 - 7) + 3.26$ = -0.66 is in excellent agreement with what it should be [log  $K = \log (6/55) = -0.96$ ]. Assuming that the affinity of methyl



**Figure 1.** Catalysed hydrolysis of methyl acetate (1 м) using (**2**) (1 mм) at pH 7.0, 25 °C.



Figure 2. pH-Rate profile for (2) (1 mM) catalysed hydrolysis of methyl acetate (1 M) at 25 °C.

acetate for aqueous copper and for (2) are comparable,  $k_2$  (3.8  $\times 10^{-1}$  s<sup>-1</sup>, half-life = 2 s) is 10<sup>9</sup> times greater than the water rate for free methyl acetate hydrolysis.<sup>7</sup> This is a spectacular rate acceleration for such a simple catalyst. Indeed, the  $k_2$  value is comparable to the  $k_{cat}$  values for chymotrypsin catalysed hydrolysis of esters<sup>11</sup> (5  $\times$  10<sup>-1</sup> s<sup>-1</sup>). However, nature's most efficient esterase that hydrolyses the neuro-transmitter, acetyl choline, is in a league by itself (acetyl choline esterase:  $k_{cat} = 3 \times 10^4$  s<sup>-1</sup>).<sup>12</sup>

$$\log K = 0.45 \,(\mathrm{p}K_{\mathrm{a}} - 7) + 3.26 \tag{2}$$

In conclusion, we have shown for the first time that Cu<sup>11</sup> can be rationally activated to catalyse the hydrolysis of a simple, unactivated ester with great efficiency.¶

¶ Detailed mechanistic analysis will be reported later in a full paper.

Financial support of this research by the Natural Sciences and Engineering Research Council of Canada is gratefully acknowledged.

Received, 13th January 1989, Com. 9/00239A

## References

- W. P. Jencks, J. Am. Chem. Soc., 1964, 86, 837; F. M. Menger and M. Ladika, *ibid.*, 1987, 109, 3145.
- 2 J. Chin and X. Zou, J. Am. Chem. Soc., 1984, 106, 3687.
- 3 J-M. Lehn and C. Sirlin, J. Chem. Soc., Chem. Commun., 1978, 949; A. J. Kirby and P. W. Lancaster, J. Chem. Soc., Perkin Trans. 2, 1972, 1206; D. J. Cram, P. Y. Lam, and S. P. Ho, J. Am. Chem. Soc., 1986, 108, 839; R. Breslow, G. Trainor, and A. Ueno, ibid., 1983, 105, 2739; D. A. Buckingham, D. M. Foster, and A. M. Sargeson, ibid., 1957, 79, 1889.
- 4 See also text books of organic, inorganic, bioinorganic, and bio-organic chemistry: A. Fersht, 'Enzyme Structure and Mechanism,' Freeman, New York, 1985, p. 67; M. N. Hughes, 'The Inorganic Chemistry of Biological Processes,' Wiley, Chichester, 1981, p. 89; M. L. Bender, R. J. Bergeron, and M. Komiyama, 'The Bioorganic Chemistry of Enzymatic Catalysis,' Wiley, New York, 1984, p. 196; J. W. Moore and R. G. Pearson, 'Kinetics and Mechanism,' Wiley, New York, 1981, p. 339; H. Dugas and C. Penny, 'Bioorganic Chemistry,' Springer-Verlag, New York, Ch. 5 and 6.
- 5 For similar mechanisms on related reactions see J. Chin, M. Banaszczyk, and V. Jubian, J. Chem. Soc., Chem. Commun., 1988, 735; J. Chin, M. Banaszczyk, V. Jubian, and X. Zou, J. Am. Chem. Soc., 1989, 111, in the press; R. L. Gustafson and A. E. Martell, J. Am. Chem. Soc., 1962, 84, 2309.
- 6 G. Anderegg, Helv. Chim. Acta, 1971, 54, 509.
- 7 J. P. Gurthrie, J. Am. Chem. Soc., 1973, 95, 6999.
- W. P. Jencks and J. Carriuolo, J. Am. Chem. Soc., 1960, 82, 1778.
  M. S. Sun and D. G. Brewer, Can. J. Chem., 1967, 45, 2729; J. Hine, 'Structural Effects on Equilibria in Organic Chemistry,' Wiley, New York, 1975, p. 244.
- 10 R. A. Cox and K. Yates, J. Am. Chem. Soc., 1978, 100, 3861.
- 11 C. Walsh, 'Enzymatic Reaction Mechanisms,' Freeman, San Francisco, 1979, p. 79.
- 12 L. Stryer, 'Biochemistry,' Freeman, New York, 1988, p. 79.